

DOI: 10.46765/2675-374X.2023V4N1P174

CONSENSUS UPDATE

## HEMOTHERAPY SUPPORT IN HSCT

Karen Lima Prata,<sup>1,2</sup> Andrea Tiemi Kondo,<sup>3</sup> Aline Miranda de Souza,<sup>4</sup> Sanny Marcelle da Costa Lira,<sup>4</sup> Andreza Alice Feitosa Ribeiro<sup>5</sup>

<sup>1</sup> Centro de Tecidos Biológicos de Minas Gerais - Fundação Hemominas.

<sup>2</sup> Agência Transfusional - Hospital das Clínicas da Universidade Federal de Minas Gerais/EBSERH.

<sup>3</sup> Departamento de Hemoterapia e Terapia Celular – Hospital Israelita Albert Einstein

<sup>4</sup> Grupo GSH.

<sup>5</sup> Unidade de Transplante de Medula Óssea do Hospital Israelita Albert Einstein.

Corresponding author: Karen Lima Prata (karenlprata@gmail.com)

Received: 16 Oct 2022 • Revised: 17 Oct 2022 • Accepted: 13 Jan 2023.

### ABSTRACT

Hemotherapy support is essential for Hematopoietic Stem Cell Transplantation (HSCT). In this article, we highlight the main points published in the 2020 SBTMO consensus and provide a brief update on the topic.

### KEY POINTS

#### MOBILIZATION AND COLLECTION OF PERIPHERAL HEMATOPOIETIC PROGENITOR CELL (HPC)

**Standard mobilization:** filgrastim (G-CSF) 10 to 20 µg/kg/day in one or two administrations for 5 days, with the first collection on day 5.<sup>(1)</sup>

**Alternative mobilization (chemotherapy and G-CSF association):** vinorelbine 35 mg/m<sup>2</sup>; cyclophosphamide (Cy) 2 to 4 grams/m<sup>2</sup> or etoposide 375 mg.<sup>(1)</sup>

**Rescue mobilization:** plerixafor (0.24 mg/kg, 9 to 12 hours before collection) with G-CSF or chemotherapy + G-CSF.<sup>(1)</sup>

**CD34<sup>+</sup> cells minimum dose:** 2 x 10<sup>6</sup>/kg per transplant (target dose of 4 to 5 x 10<sup>6</sup>/kg).<sup>(1)</sup>

**High volume apheresis** (total blood volume processed more than 4 times patient's blood volume) increases the CD34<sup>+</sup> collection.<sup>(1)</sup>

**Pediatric patients with less than 20 kg:** prime the apheresis kit with red blood cells and process at least three blood volume.<sup>(1)</sup>

#### BONE MARROW HARVEST

**Collection target:** 10-15 ml/kg of recipient, not exceeding 20 ml/kg of donor.

**Recommended cell dose:** ≥ 3x10<sup>8</sup>/kg of total nucleated cell (TNC) with minimum of 2 x 10<sup>8</sup> TCN/kg.<sup>(1)</sup>

#### PROCESSING AND CRYOPRESERVATION OF HPC-A

**Cryoprotective solutions:** Dimethyl sulfoxide (DMSO) 10% or DMSO 5% + hydroxylamine (HES) 6%, both associated with a protein source.<sup>(1)</sup>

**Final nucleated cell (NC) concentration:** 100 to 500 x 10<sup>6</sup> NC/mL.<sup>(1,2)</sup>

**Freezing ideal rate:** temperature reduction from 1 to 2° C per minute in a programmed freezing equipment or mechanical freezer.<sup>(1)</sup>

**Storage:** vapor phase or liquid nitrogen tank or mechanical freezer (-80 or -150° C).<sup>(1)</sup>

#### TRANSPORT, THAWING AND INFUSION OF HPC

**Transport of fresh products** kept the temperature between 2 and 24°C (preferably close to 4°C).<sup>(1)</sup>

**Transport of cryopreserved products:** if the cells were stored in  $-80^{\circ}\text{C}$  mechanical freezer kept the temperature at or below  $-65^{\circ}\text{C}$  (dry ice); if the cells were stored in temperature below  $-150^{\circ}\text{C}$ , kept the temperature below  $-130^{\circ}\text{C}$ .<sup>(1)</sup>

**Premedication:** hydration, diphenhydramine, dipyrone and/or hydrocortisone to prevent allergic, febrile types and/or DMSO-related reactions.<sup>(1)</sup>

**Maximum DMSO volume:** 1ml DMSO/Kg patient weight/day to reduce risk of adverse event. In lower weight pediatric patients consider removal of DMSO.<sup>(1)</sup>

**Infusion:** Do not use transfusion set device with leukocytes filter. Rate: 10 mL/minute for thawed products and 6mL/Kg of patient weight/hour for fresh products (maximum 4h).<sup>(1)</sup>

## TRANSFUSION SUPPORT

**Red blood cells (RBC), platelet and granulocyte concentrates** intended for HSCT patients should be leukoreduced and irradiated.<sup>(1)</sup>

**Platelet refractoriness:** relative common in HSCT. Causes: non-immune ( $> 80\%$ ) or immune ( $< 20\%$ ). Calculation of platelet increment is important to confirm the platelet refractoriness diagnosis.<sup>(1)</sup>

**Granulocyte transfusion** is used to prevent (patients with neutropenia or neutrophil function disorders) or treat infections in severe neutropenia (granulocytes  $< 500/\mu\text{L}$ ) patients. There are no randomized studies that prove its clinical efficacy.<sup>(1)</sup>

## ALLOGENIC BMT WITH ABO INCOMPATIBILITY

**Major or bidirectional ABO incompatibility:** RBC removal if bone marrow. Measures to reduce the anti-donor circulation isohemagglutinins (donor ABO plasma infusion or therapeutic plasmapheresis) if isohemagglutinin anti-donor  $\geq 1:32$ .<sup>(1)</sup>

**Minor ABO incompatibility:** Plasma removal if bone marrow or if HPC-A, isohemagglutinin anti-receptor  $\geq 1:256$ .<sup>(1)</sup>

**ABO blood component support:** summarized in table 1.

## DONOR LYMPHOCYTE INFUSION (DLI)

Medical evaluation of the donor is mandatory, and the eligibility criteria are the same used for blood donors.<sup>(1)</sup>

There is no need for any medication to prepare the donor for the collection.<sup>(1)</sup>

Lymphocytes can be obtained from the buffy coat of whole blood, however, the collection through apheresis equipment can offer a greater amount of CD3+ cells and is the most used.<sup>(1)</sup>

Each apheresis session should process 2 to 2.5 total blood volumes and if the number of cells needed is not obtained, a second procedure can be performed. There is a linear correlation between the number of CD3+ cells collected and the processed blood volume up to 12L.<sup>(1,3)</sup>

Dose scheme depends on the type of BMT and disease, but a staged regimen is recommended.<sup>(1)</sup>

## ANTI-HLA DONOR DESENSITIZATION

The presence of donor-specific anti-HLA antibody with mean fluorescence intensity (MFI) above 2000 is indication of desensitization protocols.<sup>(1)</sup>

## INDICATION OF PHLEBOTOMY IN IRON OVERLOAD POS BMT

Phlebotomy is a therapeutic option to drug treatment in patients with sustained hematopoiesis to reduce liver damage and irreversible tissue damage. It is indicated in cases with ferritin above  $2500\mu\text{g/L}$  and transferrin saturation close to  $100\%$ .<sup>(1)</sup>

During the revision of the chapter published in the SBTMO consensus in 2020<sup>(1)</sup>, we noticed the lack of a guidance on choosing the blood component to be transfused in the recipient undergoing transplantation with ABO incompatibility, which is now summarized in Table 1.<sup>(4)</sup>

There were few updates on the literature. We emphasize the safe nucleated cells concentration for cryopreservation, which can be up to 500,000 cells/mL after the cryoprotective solution addition.<sup>(2)</sup>

On the other hand, there have been many advances in Cell Therapy. SBTMO has published a technical manual on the topic that contains specific chapters of lymphocytes collection of by apheresis,<sup>(5)</sup> cryopreservation and transportation of mononuclear cells,<sup>(6)</sup> thawing and infusion of CAR-T cells,<sup>(7)</sup> among others.

At the national level, the current health legislation was updated.<sup>(8)</sup>

**TABLE 1.** Recommendation for post-transplantation transfusion support in ABO-incompatible HPC recipients.(4)

Receptor	Donor	Incompatibility type	Fase I	Fase II			Fase III	
			All blood components	RBC concentrate	Platelet concentrate		Plasma	All blood components
					1st choice	Other choice		
A	O	Minor	A	O	A	AB; B; O	A; AB	O
B	O	Minor	B	O	B	AB; A; O	B; AB	O
AB	O	Minor	AB	O	AB	A; B; O	AB	O
AB	A	Minor	AB	A	AB	A; B; O	AB	A
AB	B	Minor	AB	B	AB	B; A; O	AB	B
O	A	Major	O	O	A	AB; B; O	A; AB	A
O	B	Major	O	O	B	AB; A; O	B; AB	B
O	AB	Major	O	O	AB	A; B; O	AB	AB
A	AB	Major	A	A	AB	A; B; O	AB	AB
B	AB	Major	B	B	AB	B; A; O	AB	AB
A	B	Bidirecional	A	O	AB	A; B; O	AB	B
B	A	Bidirecional	B	O	AB	B; A; O	AB	A

**Phase I:** from the beginning of patient preparation to the beginning of conditioning; **Phase II:** from the beginning of conditioning until the direct antiglobulin test becomes negative and the direct typing of the patient becomes the same as the donor; **Phase III:** from the moment when the direct and reverse typing of the patient is the same as the donor.

**REFERENCES**

1. Kondo AT, Prata KL, Souza AM, et al. Transfusion support in HSCT; JBMTCT. 2021;2(1):235-44.
2. Belisario AR, Costa AP, Luz JR, et al. Influence of laboratory procedures on postthawing cell viability and hematopoietic engraftment after autologous peripheral blood stem cell transplantation. Transfusion. 2021;61(4):1202-14.
3. Sato H, Shiobara S, Yasue S, et al. Lymphocyte collection for donor leucocyte infusion from normal donors: estimation of the minimum processed blood volume and safety of the procedure. Vox Sang. 2001;81(2):124-7.
4. O'Donghaile D, Kelley W, Klein HG, et al. Recommendations for transfusion in ABO-incompatible hematopoietic stem cell transplantation. Transfusion. 2012;52(2):456-8.
5. Souza AM. Collection of lymphocytes by apheresis. JBMTCT. 2022;3(1):26-34.
6. Prata KL, Kondo AT. Peripheral blood mononuclear cells cryopreservation and transportation: PBMC cryopreservation and transportation. JBMTCT. 2022;3(1):45-56.
7. Kondo AT, Kerbauy LN, Alvarez KC, et al. Thawing and infusion of CAR-T cell products. JBMTCT. 2022;3(1):63-9.
8. Anvisa. Resolução RDC no 508, de 27 de maio de 2021. Dispõe sobre as boas práticas em células humanas para uso terapêutico e pesquisa clínica e dá outras providências [Internet]. Brasília; 2021 [cited 2023 Jan 23]. Available from: <https://www.in.gov.br/en/web/dou/-/resolucao-rdc-n-508-de-27-de-maio-de-2021-323013606>